

ANTIPHOSPHOLIPID ANTIBODIES IN HEALTHY SERBIAN MIDDLE-AGED SUBJECTS: PRELIMINARY DATAANTIFOSFOLIPIDNA ANTITELA U ZDRAVIH SRPSKIH OSOBA SREDNJIH GODINA:
PRELIMINARNI PODACIMirjana B. Bećarević^{1*}, Snežana Jovičić^{2,3}, Svetlana D. Ignjatović^{2,3}, Duško Mirković^{2,3}¹University of Novi Sad, Faculty of Medicine, Department of Pharmacy, Novi Sad, Serbia²University of Belgrade, Faculty of Pharmacy, Department of Medical Biochemistry, Belgrade, Serbia³Center for Medical Biochemistry, Clinical University Center of Serbia, Belgrade, Serbia**Summary**

Background: The investigation of the prevalence of the IgG and the IgM isotypes of anticardiolipin (aCL) and anti- β 2glycoprotein I (a β 2gpl) Abs in healthy Serbian middle-aged subjects was the main goal of our study. In addition, we analyzed the potential associations of above-mentioned Abs with serum proteins and lipids/lipoproteins.

Methods: Forty healthy subjects were included in our study. Obesity (BMI ≥ 30 kg/m²) was present in 8/40 (20%) subjects. Titers of analyzed Abs were measured by ELISA.

Results: The prevalence of IgG and IgM a β 2gpl Abs was 5% and 12.5%, respectively, while the prevalence of IgM aCL was 10%. The IgG a β 2gpl Abs were significantly different between subjects with normal triglycerides levels and those with hypertriglyceridemia (Mann-Whitney, $P = 0.014$). The significant difference in hsCRP concentrations was observed between subjects with the increased levels of the IgM isotype of aCL Abs and those with normal IgM aCL values (Mann-Whitney, $P = 0.028$).

Conclusions: Dyslipidemia and BMI ≥ 30 were associated with aPL Abs and therefore, the correction of BMI and lipid status might be beneficial in reduction or elimination of predisposing factors that might trigger thrombotic events in otherwise healthy middle-aged subjects. Larger national study is necessary to confirm our findings.

Keywords: antiphospholipid antibodies, apolipoproteins, complement components, C-reactive protein, haptoglobin, serum amyloid A

Kratka sadržaj

Uvod: Analiza prevalentnosti IgG i IgM izotipa antikardiolipinskih (aCL) i anti- β 2glikoprotein I (a β 2gpl) At kod zdravih sredovećnih stanovnika Srbije je bila glavni cilj naše studije. Dodatno, analizirali smo potencijalnu povezanost gorenavedenih At sa serumskim proteinima i lipidima/lipoproteinima.

Metode: 40 zdravih ispitanika je bilo uključeno u našu studiju. Gojaznost (BMI ≥ 30 kg/m²) je uočena kod 8/40 (20%) osoba. Titri analiziranih antitela su utvrđivani ELISA testom.

Rezultati: Prevalentnost IgG i IgM a β 2gpl At je bila 5% i 12.5%, redom, dok je prevalentnost IgM aCL bila 10%. Nivoi IgG a β 2gpl At su se značajno razlikovali između ispitanika sa i bez hipertrigliceridemije (Mann-Whitney, $P = 0.014$). Značajne razlike u hsCRP koncentracijama uočene su između osoba sa povišenim nivoima IgM aCL At i onih sa referentnim vrednostima (Mann-Whitney, $P = 0,028$).

Zaključak: Dislipidemija i BMI ≥ 30 su bili povezani sa aPL At uprkos njihovoj niskoj prevalentnosti, i zato korekcija BMI i lipidnog statusa bi bila korisna u redukciji ili eliminaciji predispozirajućih faktora koji mogu da izazovu trombotički događaj kod inače zdravih sredovećnih ispitanika. Obimnije nacionalne studije su neophodne da bi potvrdile naše nalaze.

Ključne reči: antifosfolipidna antitela, apolipoproteini, komplementne komponente, C-reaktivni protein, haptoglobin, serum amilod A

Address for correspondence:

Professor Mirjana Bećarević, PhD
University of Novi Sad, Faculty of Medicine,
Department of Pharmacy, Novi Sad, Serbia
Hajduk Veljkova 3, 21000 Novi Sad, Serbia
e-mail: bm48mb32@gmail.com and
mirjana.becarevic@mf.uns.ac.rs
ORCID: 0000-0003-2114-8621

List of abbreviations: Abs, antibodies; a β 2gpl, anti- β 2glycoprotein I Abs; aCL, anticardiolipin Abs; aPL, antiphospholipid Abs; APS, antiphospholipid syndrome.

Introduction

Antiphospholipid antibodies (aPL Abs) represent a diverse group of Abs directed against complexes formed between negatively charged phospholipids (i.e. cardiolipin (CL), phosphatidyl-serine, phosphatidyl-inositol etc.) or blood proteins (i.e. beta 2 glycoprotein I ($\beta 2$ gpl)), etc (1–5).

Increased titers of aPL Abs are the main laboratory feature of the antiphospholipid syndrome (APS). It is an autoimmune disease that is (beside the presence of aPL Abs) characterized by the presence of recurrent thrombosis and/or pregnancy losses (6, 7). According to the latest classification criteria for the diagnosis of the APS (8, 9), the presence of aPL Abs (i.e. the IgG and/or the IgM isotype of the anticardiolipin (aCL) and/or the IgG and/or the IgM isotype of the anti- $\beta 2$ glycoprotein I ($\beta 2$ gpl) Abs) must be present at medium to high titers in two or more occasions, at least twelve weeks apart. The reasons why $\beta 2$ gpl and CL, as ubiquitously present autoantigens in some persons promotes production of pathogenic autoAbs remains elusive.

There are several studies that investigated the prevalence of aPL Abs in healthy subjects of various nationalities (10–12) and numerous studies that compared levels of aPL Abs in patients with different autoimmune disease vs. healthy subjects (13–16). We have previously reported (13) that in 47 Serbian young (mean \pm SD, 39.68 ± 13.93 , 33 female) lean, healthy adults, levels of aPL Abs were significantly lower in comparison to patients with primary APS and that levels of analyzed serum lipids (cholesterol, HDL-cholesterol, LDL-cholesterol and triglycerides) were below cut-off values in these young subjects. However, no studies that analyze the association of aPL Abs with sera lipids/lipoproteins and proteins in healthy Serbian middle-aged subjects are available. Therefore, the aim of our study was to evaluate the prevalence of aPL Abs in healthy middle-aged Serbian subjects and to investigate the potential correlation between aPL Abs positivity and lipids/lipoproteins (cholesterol, HDL-cholesterol, LDL-cholesterol, triglycerides, apolipoproteins (apo) A1, B and lipoprotein (a) (Lp(a)) and serum proteins (C-reactive protein (CRP), serum amyloid A (SAA), haptoglobin (HPT), fibrinogen, C3 and C4 complement components).

Materials and Methods

Subjects

All procedures performed in our study were in accordance with Helsinki declaration (and its later amendments) and with the ethical standards of the institutional ethical committee. Written informed consent was obtained from all individual participants included in the study.

Our study included 40 healthy subjects (mean age \pm SD, 58.22 ± 3.47) comprised from our colleagues and our acquaintances that did not show any clinical signs of thrombosis, pregnancy morbidity, infections, cancer and autoimmune diseases. The use of the laboratory information system data for this study was approved by our local Ethical Committee (Ethical Committee of the Clinical University Center of Serbia, Approval No 4815/3). Female to male ratio was 16/24. The body mass index (BMI) was calculated as the weight (kg)/height² (m²).

Methods

Serum concentrations of total cholesterol (TC), HDL-cholesterol (HDL-C), LDL-cholesterol (LDL-C), triglycerides (TG) and high sensitivity C-reactive protein (hsCRP) were measured on Olympus AU2700 automated analyzer (Beckman Coulter Inc, USA). Total cholesterol and TG concentrations were determined using standard enzymatic assays. High-density lipoprotein cholesterol concentrations were determined using direct enzymatic method. Friedwald formula was used to calculate LDL-C concentrations, but if TG concentrations >4.50 mmol/L direct enzymatic method was used.

Concentrations of hsCRP were measured with an immunoturbidimetric latex assay. Cut-off values for TC, HDL-C, LDL-C, TG and hsCRP were 5.0 mmol/L, 1.55 mmol/L, 2.5 mmol/L, 1.70 mmol/L, 3 mg/L (for high cardiovascular risk), respectively.

Apolipoprotein A-I, apoB, Lp(a), C3 and C4 complement components (C3, C4) were determined by immunoturbidimetric procedures on Architect c8000 chemistry system (Abbott Laboratories, Illinois, USA). Reference ranges for apoA1 and apoB were: 0.95–1.86 g/L (men), 1.01–2.23 g/L (women); 0.49–1.73 g/L (men), 0.53–1.82 g/L (women), respectively. Cut-off value for Lp(a) was 300 mg/L. Reference ranges for C3 and C4 complement components were: 0.82–1.85 g/L (men), 0.83–1.93 g/L (women); 0.15–0.53 g/L (men), 0.15–0.57 g/L (women), respectively (17).

Serum concentration of SAA and HPT were assayed using particle-enhanced immunonephelometry with BN II nephelometer (Siemens Healthcare GmbH, Germany). Cut-off value for SAA was 6.4 mg/L, while the reference range for HPT was 0.3–2 g/L. Fibrinogen concentrations were measured in citrate plasma by prothrombin time (PT)-based method on ACL 7000 analyzer (Instrumentation Laboratory SpA, Milan, Italy). For each analyzer appropriate supplied reagents were used. Reference range for fibrinogen was 1.7–5.4 g/L.

Antibody levels were estimated by ELISA in patient sera using commercially available reagents of ORGENTEC, Diagnostika GmbH, Germany for the detection of anti- $\beta 2$ gpl (IgG and IgM isotypes) and

anticardiolipin (aCL) (IgG and IgM isotypes) antibodies. Cut-off values were set in accordance to manufacturer recommendation (8 U/mL (for the IgG and IgM isotypes of aβ2gpl Abs), 10 GPL-U/mL (for IgG aCL) and 7 MPLU/mL (for IgM aCL Abs).

Statistical analysis

Shapiro-Wilk test was used to study whether analyzed variables followed a normal distribution. The categoric variables were expressed in percentages (%), while continuous variables were expressed as mean ± SD in the case of normal distribution, but if concentrations did not follow a normal distribution pattern, the values were expressed as median (25th - 75th percentiles). Mann-Whitney test, Kruskal-Wallis and 2-test were used, when appropriate. The correlation between two quantitative variables was determined with the Spearman's correlation test. In all of the above-mentioned tests, $P < 0.05$ was considered statistically significant. Analyses were conducted in SPSS 20 (SPSS, Inc, Chicago, IL, USA).

Results

Obesity (BMI ≥ 30 kg/m²) was present in 2/16 (12.5%) female subjects and in 6/24 (25%) male subjects. Increased BMI values (BMI 25–30 kg/m²) were present in eight (50%) female and in eight (33.33%) male subjects. Our study included 5/16 (31.25%) female smokers and 6/24 (25%) male smokers.

Serologic features of analyzed subjects are presented in *Table 1*. Not a single subject had increased titers of the IgG isotype of aCL Abs. Increased levels of the IgM isotype of aCL Abs were present in 4 (2 female and 2 male)/40 (10%) of analyzed subjects. Elevated levels of the IgG isotype of aβ2gpl Abs were present in only two male subjects (5%), while increased IgM aβ2gpl levels were observed in 5 (4 female and 1 male)/40 (12.5%) of analyzed subjects. Simultaneous presence of the IgM isotype of both aCL and aβ2gpl Abs was present in three subjects (7.5%).

The IgM isotype of aCL and aβ2gpl Abs were in positive correlation ($r = 0.882$, $P = 0.000$) (*Figure 1*,

Table 1 Concentrations analyzed parameters in female and male subjects (comparison was done by Mann-Whitney, * $P < 0.05$).

Parameters Median (25th–75th)	Female n = 16	Male n = 24	P value
Age (years, (mean ± SD))	57.19 ± 3.31	58.92 ± 3.48	0.130
BMI (kg/m ²)	26.34 (24.33 – 28.86)	27.44 (23.77 – 30.04)	0.782
Glucose (mmol/L)	5.25 (5.02 – 6.02)	5.45 (5.20 – 5.87)	0.589
Cholesterol (mmol/L)	6.23 (5.80 – 7.60)	6.15 (5.69 – 6.77)	0.464
Triglycerides (mmol/L)	1.63 (1.36 – 2.39)	1.57 (1.06 – 2.61)	0.879
LDL-cholesterol (mmol/L)	4.23 (3.71 – 4.84)	3.78 (3.15 – 4.30)	0.100
HDL-cholesterol (mmol/L)	1.30 (1.15 – 1.71)	1.28 (1.05 – 1.48)	0.499
ApoAI (g/L)	1.58 (1.47 – 1.75)	1.56 (1.38 – 1.77)	0.629
ApoB (g/L)	1.24 (1.09 – 1.44)	1.25 (1.05 – 1.42)	0.782
Lp(a) (g/L)	0.17 (0.08 – 0.55)	0.16 (0.03 – 0.32)	0.362
CRP (mg/L)	2.05 (1.29 – 3.76)	1.54 (1.08 – 3.37)	0.508
SAA (mg/L)	4.55 (2.22 – 9.05)	3.35 (1.45 – 6.85)	0.334
Fibrinogen (g/L)	3.91 (2.97 – 4.50)	4.22 (3.8 – 4.47)	0.163
C3 (g/L)	1.43 (1.30 – 1.51)	1.51 (1.13 – 1.63)	0.659
C4 (g/L)	0.27 (0.24 – 0.32)	0.26 (0.21 – 0.30)	0.415
HPT (g/L)	1.39 (0.98 – 1.67)	1.21 (0.98 – 1.72)	0.730
aCL IgG (GPL U/mL)	3.29 (1.89 – 4.33)	2.84 (1.59 – 3.55)	0.282
aCL IgM (MPL U/mL)*	2.76 (1.79 – 5.07)	2.03 (1.33 – 2.78)	0.034*
ab2gpl IgG (U/mL)	2.54 (1.77 – 3.44)	2.16 (1.66 – 2.77)	0.281
ab2gpl IgM (U/mL)*	3.63 (2.19 – 7.86)	2.19 (1.47 – 3.55)	0.011*

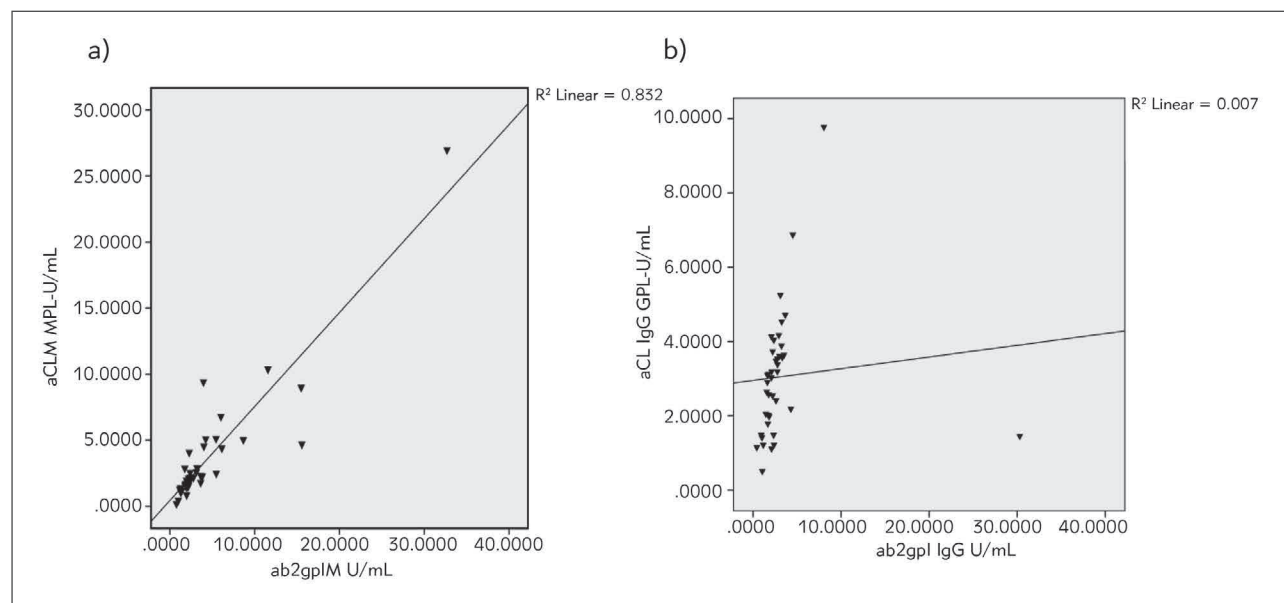


Figure 1 Correlation between the IgM ($r = 0.882$, $P = 0.000$, (Panel (a) and the IgG ($r = 0.632$, $P = 0.000$, (Panel (b) isotypes of anticardiolipin (aCL) and anti- β 2glycoprotein I (ab2gpl) antibodies

Panel A). The IgM class of aCL Abs and the IgG isotype of β 2gpl Abs were in positive correlation ($r = 0.319$, $P = 0.045$). A positive correlation was found for the IgG and the IgM isotype of aCL Abs ($r = 0.502$, $P = 0.001$). In addition, the IgG isotype of aCL Abs was in positive correlation with both the IgG ($r = 0.632$, $P = 0.000$) (Figure 1, Panel B) and the IgM ($r = 0.421$, $P = 0.007$) isotypes of β 2gpl antibodies.

The hypertriglyceridemia was noticed in 17/40 (42.5%) of analyzed subjects and the concentrations of the IgG isotype of β 2gpl Abs were significantly different between subjects with and without hypertriglyceridemia (Mann-Whitney, $P = 0.014$). Subjects with increased Lp(a) levels and those with normal Lp(a) values had significantly different IgG β 2gpl Abs concentrations (Mann-Whitney, $P = 0.026$). However, only male subjects showed a positive correlation between Lp(a) and the IgM isotype of β 2gpl Abs ($r = 0.412$, $P = 0.045$).

Kruskal-Wallis test (One-way ANOVA) revealed that the IgG isotype of β 2gpl Abs ($P = 0.020$), hsCRP ($P = 0.048$), C3c ($P = 0.015$), cholesterol ($P = 0.028$) and triglycerides ($P = 0.012$) concentrations were significantly different among subjects with different BMI values (obese, increased and normal BMI). The significant difference in body weight values was observed between subgroups of subjects with increased IgM isotype of β 2gpl Abs and those without it (i.e. normal IgM β 2gpl concentrations, Mann-Whitney, $P = 0.034$), while significant difference in hsCRP concentrations was observed between subjects with the increased levels of the IgM isotype of aCL Abs and those with normal IgM aCL values (Mann-Whitney, $P = 0.028$).

No correlation between the analyzed Abs and C3c and C4 complement components, SAA, CRP, HPT, fibrinogen and apolipoproteins was obtained. However, hsCRP was in positive correlation with SAA ($r = 0.511$, $P = 0.001$), C3c ($r = 0.469$, $P = 0.002$), C4 ($r = 0.377$, $P = 0.017$) and HPT ($r = 0.388$, $P = 0.013$). Fibrinogen concentrations were in positive correlation with triglycerides ($r = 0.332$, $P = 0.037$) and C3c ($r = 0.479$, $P = 0.002$).

Discussion

McIntyre et al (12) have published that 63/775 (8.1%) volunteer blood donors (average age: 43 years, range (17–82)) had positive finding of one or more aPL Abs (12). Another study (18) has revealed that 20.7% of centenarians were positive for IgG aCL and 2.59% for IgM aCL Abs, while 54.3% of centenarians were positive for the IgG β 2gpl and 8.6% for IgM β 2gpl Abs (18). Despite this high aPL Abs prevalence in centenarians (comparable to titers observed in APS patients), the authors of the study did not observe a single vascular event and therefore the authors have suggested that some »unknown protective factor and/or lacking of triggering factors« are responsible for their findings (18). Similarly, Mustonen et al. (19) have reported that single aPL Abs positivity does not seem to carry an elevated risk of thrombosis (in asymptomatic aPL Abs carriers). Avčín et al. (20) reported no statistically significant differences in the frequency of the elevated either aCL isotype between blood donors (mean age: 34 years, range (18 – 65)) and analyzed children (preschool and adolescent), i.e. 5/52 (9.6%) blood donors were positive for aCL Abs (5.8% were positive for IgG aCL

Abs vs. 7/61 (11.4%) of analyzed children were positive for IgG aCL). The same group (20) have reported no differences in the frequency of the either isotype of a β 2gpl Abs between blood donors and analyzed children (4/52 (7.7%) of blood donors were positive for a β 2gpl Abs and 1/52 (1.9%) were positive for IgG a β 2gpl Abs) vs. 4/61 (6.6%) of analyzed children and (2/61 (3.3% were positive for IgG aB2gpl Abs)). However, Avčin et al. (20) did not analyze the gender differences in regard with the prevalence of aPL Abs in blood donors and children. McIntyre et al. (12) have reported that males were positive for aCL more often than females, but no differences in aPL isotypes between gender was observed (12). In addition, the same authors (12) have reported the persistence of the IgM aCL Abs (in a healthy male repeat blood donor (age: 60 years)) even after 16 months interval between blood draws. Although one might expect the isotype switch from IgM to IgG to occur with the loss of the IgM isotype in the normal antibody response, the authors (12) did not provide an explanation for their observation.

In comparison to above-mentioned studies, our study population was comprised from middle-aged subjects (mean age \pm SD, 58.22 \pm 3.47) and we observed that not a single subject had increased titers of the IgG isotype of aCL Abs, but elevated IgM aCL Abs levels were present in 10% of analyzed persons, while increased titers of the IgG and IgM a β 2gpl were observed in 5% and 12.5% of analyzed subjects, respectively. We have observed that the IgM isotype was more frequent than the IgG isotype but this was not statistically significant.

Previously it was reported that β 2gpl (also known as apolipoprotein (apo) H) was implicated in atherogenic and thrombotic processes and that serum β 2gpl concentrations were elevated in patients with primary hyperlipidaemia (21). In addition, significant correlations were observed between β 2gpl and triglycerides and total cholesterol concentrations (21). Although in our study we did not measure serum β 2gpl levels, we observed that IgG a β 2gpl Abs concentrations were significantly different between subjects with and without hypertriglyceridemia. In addition, the concentrations of IgG a β 2gpl Abs were significantly different between subjects with increased Lp(a) levels and those with normal levels. Interestingly, only analyzed male subjects showed a positive correlation between the IgM a β 2gpl Abs and Lp(a) concentrations.

Previously it was reported that concentrations of IgG and IgM aCL Abs were similar in obese and nonobese patients with primary APS (22). In agreement, no differences in regard to either isotype of aCL Abs were observed in our subjects considering their BMI values. However, in our group, obesity was present in 12.5% of females and in 25% of males and we noticed that subjects with different BMI values had

significantly different concentrations of the IgG a β 2gpl Abs, hsCRP and C3c. Elevated hsCRP levels and the presence of aPL Abs exhibited some similarities in the pathogenesis of thrombosis (23). It is considered that hsCRP is a predictor of vascular events independently of all other lipids and non-lipid risk factors (23). Lin et al. (24) have reported that in patients with inflammation, β 2gpl levels were in negative correlation with CRP and in positive correlation with negative acute phase proteins (such as albumin and transferrin). It was reported that acute phase proteins (such as SAA) have been associated with the pathology of anti- β 2gpl Abs and that SAA levels were increased and correlated with the history of thrombosis in APS patients (25), while in healthy young Japanese, no correlation between CRP and SAA levels was observed (26). In our study, hsCRP concentrations were significantly different between subjects with increased IgM aCL Abs titers and those without it. In addition, we observed significant correlation between hsCRP and SAA, HPT and complement components (C3c and C4). In patients with idiopathic aPL Abs, fibrinogen concentrations correlated with the aCL IgG Abs and the authors (27) suggested that measurement of fibrinogen may be beneficial in defining aPL subjects with higher thrombotic risk that might require pharmacological intervention for lowering fibrinogen levels (27). In our study, fibrinogen concentrations did not correlate with the either isotype of analyzed Abs.

In conclusion, BMI 30 (obesity) and dyslipidemia were associated with aPL Abs despite their low prevalence in analyzed subjects. Antiphospholipid antibodies are regarded as natural autoantibodies and due to »molecular mimicry between microbial epitopes and human β 2gpl it is possible that in genetically predisposed subjects, generation of aPL Abs might be initiated« (28). However, predisposing factors are not completely elucidated yet and there are several reports that suggest that pro-inflammatory cytokines and acute phase reactants (25) are important for generating »second hit« that is vital in the pathology of aPL Abs. Our study provides a rationale for the fact that correction of BMI and lipid status might be beneficial in reduction and/or elimination of predisposing factors that might trigger thrombotic events in otherwise healthy Serbian middle-aged subjects. In addition, our results should be regarded with caution (i.e. as preliminary) due to relatively small number of participants included in the study and therefore, larger national study is necessary to confirm our findings.

Acknowledgment

The authors of the article have no conflict of interest (financial nor non-financial) related to this manuscript. All authors approved the final manuscript as submitted and agree to be accountable for all

aspects of the work (MB is responsible for analysis, interpretation of data and writing of the Article; SJ and MB were responsible for laboratory measurements of investigated parameters; SJ was responsible for recruitment of subject and providing their data; SI and DM are responsible for the final approval of the Article).

The present work was supported by the Ministry of Science, Education and Technological Develop-

ment of the Republic of Serbia on the basis of contracts No.175036 and No.451-03-68/2020-14/200161.

Conflict of interest statement

All the authors declare that they have no conflict of interest in this work.

References

- Alessandri C, Conti F, Pendolino M, Mancini R, Valesini G. New autoantigens in the antiphospholipid syndrome. *Autoimmunity Rev* 2011; 10: 609–16.
- Bećarević M. The IgG and IgM isotypes of anti-annexin A5 antibodies: relevance for primary antiphospholipid syndrome. *J Thromb Thrombolysis* 2016; 42: 552–7.
- Bećarević M. Antibodies against complement components: relevance for the antiphospholipid syndrome-biomarkers of the disease and biopharmaceuticals. *Curr Rheumatol Rep* 2017; 19: 1–9. DOI 10.1007/s11926-017-0669-1.
- Bećarević M. Detrimental roles of TNF-alpha in the antiphospholipid syndrome and de novo synthesis of antiphospholipid antibodies induced by biopharmaceuticals against TNF-alpha. *J Thromb Thrombolysis* 2017; 44: 565–70.
- Bećarević M, Stojanovich Lj, Ignjatović S, Dopsaj V. The IgM Isotype of Anti-Annexin A5 Antibodies and Multiple Positivity of Conventional Antiphospholipid Antibodies: Increasing the Number of Clinical Manifestations of Primary Antiphospholipid Syndrome. *Clin Rheum*. 2016; 35: 1361–1365, DOI 10.1007/s10067-016-3230-0. issn: 0770-3198.
- Bertolaccini ML, Amengual O, Andreoli L, et al. 14th International Congress on Antiphospholipid Antibodies Task Force. Report on antiphospholipid syndrome laboratory diagnostics and trends. *Autoimmun Rev* 2014; 13: 917–30.
- Bećarević M, Mirković D, Ignjatović S. Double positivity of the IgG isotype of both anticardiolipin and anti-b2gpl antibodies is associated with the highest number of vascular impairment parameters in patients with primary antiphospholipid syndrome: preliminary data. *Clin Rheum* 2016: DOI 10.1007/s10067-016-3438-z
- Miyakis S, Lockshin MD, Atsumi T, et al. International consensus statement on an update of the classification criteria for definite antiphospholipid syndrome. *J Thromb Haemost* 2006; 4: 295–306.
- Sciascia S, Radin M, Cecchi I, Levy RA, Erkan D. 16th International congress on antiphospholipid antibodies task force report on clinical manifestations of antiphospholipid syndrome. *Lupus* 2021; 30(8) 1314–26.
- Montaruli B, De Luna E, Mengozzi G, et al. Anti-cardiolipin and anti-b2-glycoprotein I antibodies: normal reference ranges in northwestern Italy. *Lupus* 2012; 21: 799–801.
- Martínez-Flores JA, Serrano M, Pérez D, Lora D, Paz-Artal E, Morales JM, Serrano A. Detection of circulating immune complexes of human IgA and beta 2 glycoprotein I in patients with antiphospholipid syndrome symptomatology. *J Immunol Methods* 2015; DOI 10.1016/j.jim.2015.04.002.
- McIntyre JA, Wagenknecht DR, Waxman DW. Frequency and specificity of antiphospholipid antibodies (aPL) in volunteer blood donors. *Immunobiol* 2003; 207: 59–63.
- Bećarević M, Andrejević S, Miljić P, Bonači-Nikolić B, Majkić-Singh N. Serum lipids and anti-oxidized LDL antibodies in primary antiphospholipid syndrome. *Clin Exp Rheumatol* 2007; 25 (3): 361–6.
- Bećarević M, Čabarkapa V, Đerić M, Ignjatović S. Antiphospholipid antibodies and renal impairment parameters in diabetic nephropathy: preliminary data. *Clin Appl Thromb Hemost* 2016; DOI 10.1177/1076029616642512.
- Bećarević M, Sarić Matutinović M, Žarković M, Nedeljković Beleslin B, Ćirić J, Ignjatović S. Antiphospholipid Antibodies in Patients with Graves' orbitopathy: Preliminary Data *Endocrine* 2021; DOI 10.1007/s12020-021-02769-z.
- Ghafar MTA, El-Masry MI. Verification of quantitative analytical methods in medical laboratories. *J Med Biochem* 2021; 40 (3): 225–36.
- Bećarević M, Ignjatović S. Proinflammatory proteins in female and male patients with primary antiphospholipid syndrome: preliminary data. *Clin Rheumatol* 2016; 35(10): 2477–83. DOI 10.1007/s10067-016-3345-3.
- Meroni PL, Mari D, Monti D, Coppola R, Capri M, Salvioli S, Tincani A, Gerli R, Franceschi C. Antibeta 2 glycoprotein I antibodies in centenarians. *Experimental Gerontology* 2004; 39: 1459–65.
- Mustonen, P, Lehtonen KV, Javela K, Puurunen M. Persistent antiphospholipid antibody (aPL) in asymptomatic carriers as a risk factor for future thrombotic events: a nationwide prospective study. *Lupus* 2014; 23: 1468–76.
- Avčin T, Ambrožič A, Kuhar M, Kveder T, Rozman B. Anticardiolipin and anti-b2glycoprotein I antibodies in

- sera of 61 apparently healthy children at regular preventive visits. *Rheumatol* 2001; 40: 565–73.
21. Crook MA. Apolipoprotein H: its relevance to cardiovascular disease. *Atherosclerosis* 2010; 209: 32–4.
 22. Caldas CA, da Mota LMH, de Carvalho JF. Obesity in primary antiphospholipid syndrome is associated with worse outcome. *Joint Bone Spine* 2011; 78: 319–25.
 23. Miesbach W, Gokpınar B, Gilzinger A, Claus D, Scharrer I. Predictive role of hs-C-reactive protein in patients with antiphospholipid syndrome. *Immunobiol* 2005; 210: 755–60.
 24. Lin F, Murphy R, White B, et al. Circulating levels of B2gpl in thrombotic disorders and in inflammation. *Lupus* 2006; 15: 87–93.
 25. Artenjak A, Omersel J, Ahlin Grabnar P, et al. Oxidatively altered IgG with increased immunoreactivity to b2-glycoprotein I and its peptide clusters influence human coronary artery endothelial cells. *Lupus* 2015; 24: 448–62.
 26. Kokubun M, Imafuku Y, Okada M, et al. Serum amyloid A (SAA) concentrations varies among rheumatoid arthritis patients estimated by SAA/CRP ratio. *Clin Chim Acta* 2005; 36: 97–102.
 27. Ames PRJ, Margarita A, Delgado Alves J, Tommasino C, Iannaccone L, Brancaccio V. Anticardiolipin antibody titre and plasma homocysteine level independently predict intima media thickness of carotid arteries in subjects with idiopathic antiphospholipid antibodies. *Lupus* 2002; 11: 208–14.
 28. van den Hoogen LL, van Roon JAG, Radstake TRDJ, Fritsch-Stork RDE, Derksen RHWM. Delineating the deranged immune system in the antiphospholipid syndrome. *Autoimmunity Rev* 2015; DOI 10.1016/j.autrev.2015.08.011.

Received: January 10, 2022

Accepted: January 24, 2022